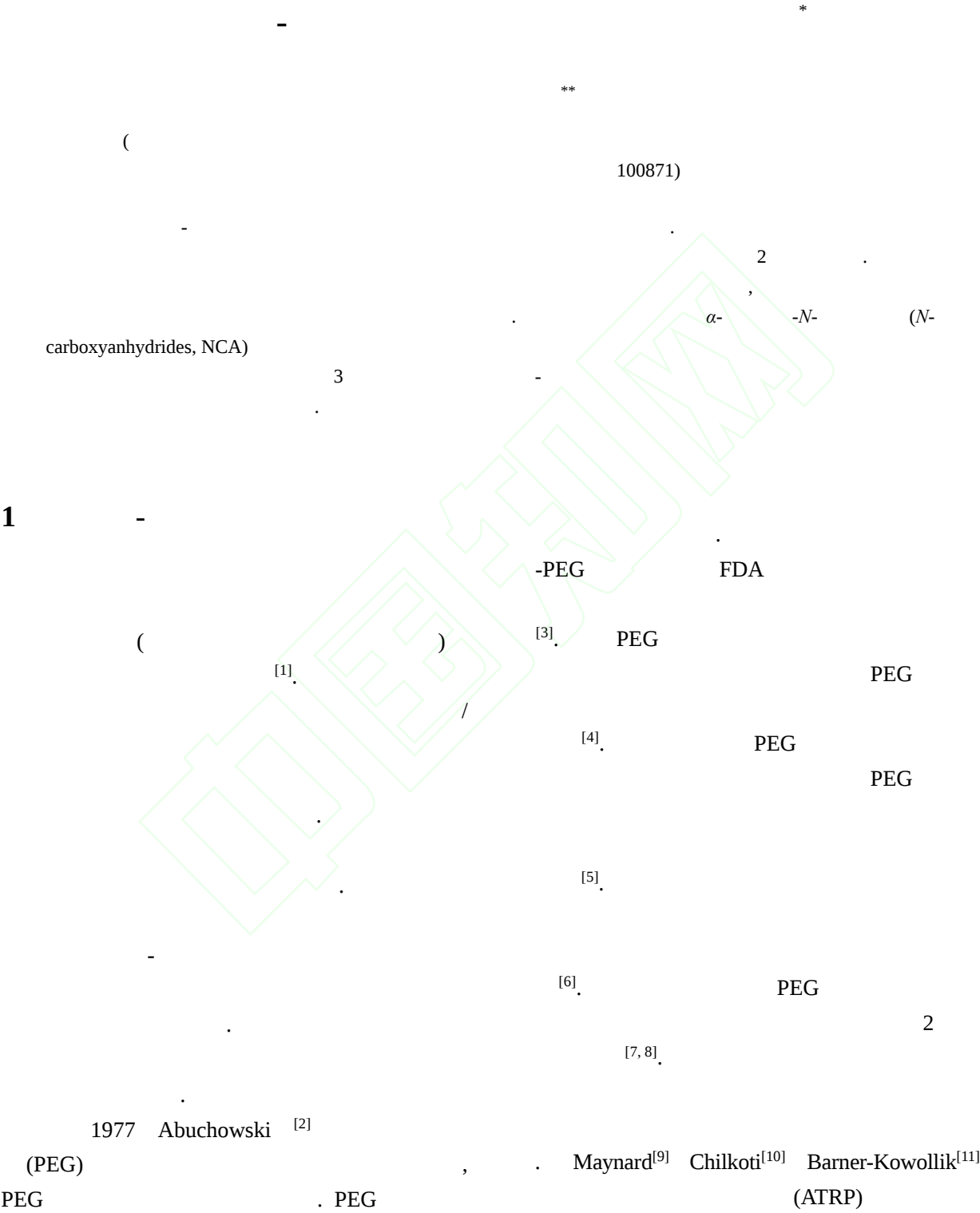


· 专论 ·

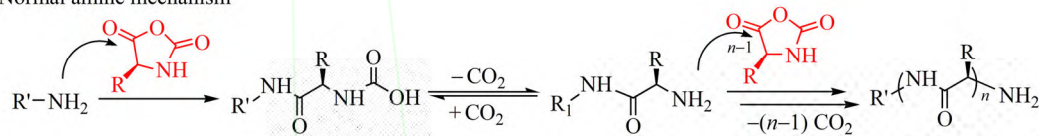


* 2017-07-28 2017-09-22 (21474004, 21434008)
** E-mail: chemhualu@pku.edu.cn
doi: 10.11777/j.issn1000-3304.2018.17204

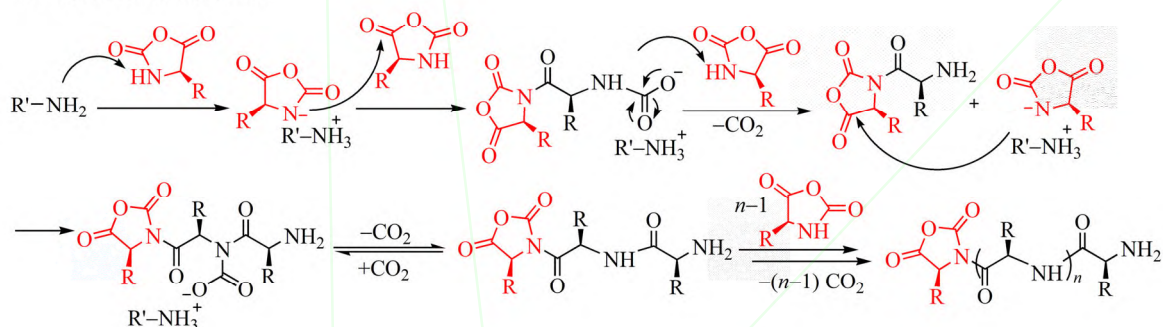
- (RAFT)
l grafting-from L

[12] α - (IFN- α) C ATRP

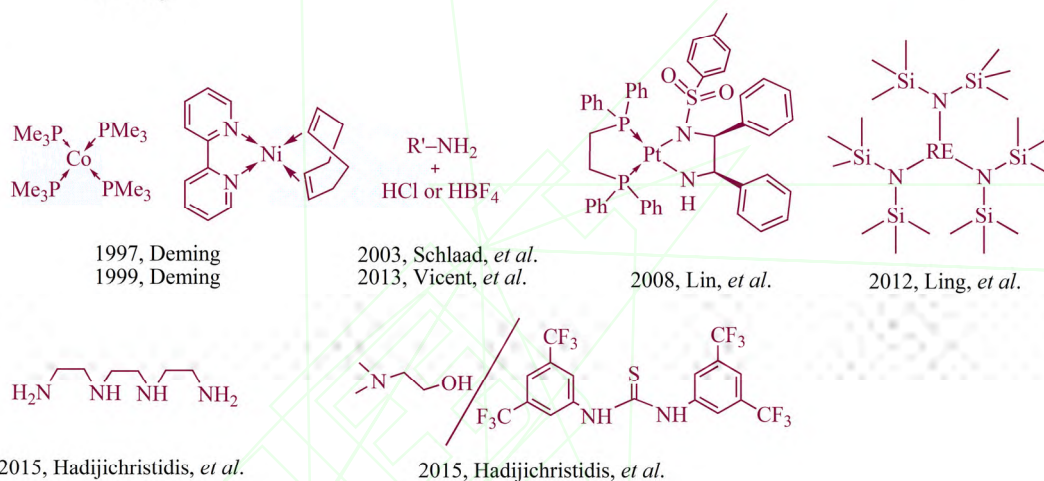
(a) Normal amine mechanism



Active monomer mechanism

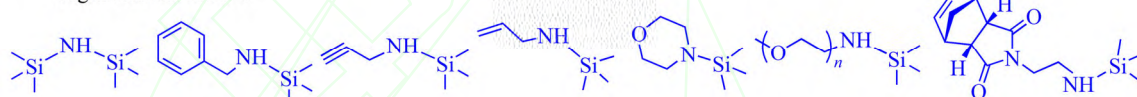


(b)

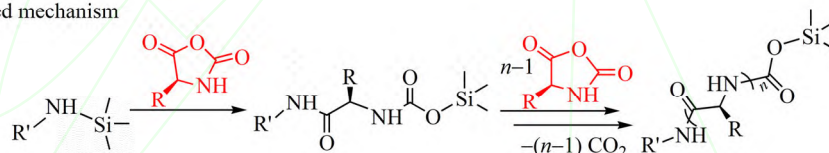


(c)

Organosilicon amines



Proposed mechanism



(d)

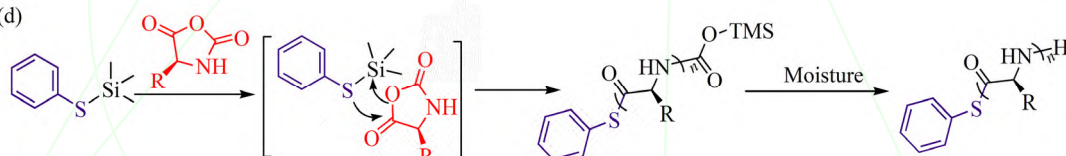


Fig. 1 (a) Two general ring-opening polymerization (ROP) mechanisms of α -amino acid NCAs; (b) Examples of reported initiation systems successfully used in controlled ROP of NCAs; (c) Structures of organosilicon amines and proposed mechanisms; (d) PhS-TMS mediated NCA polymerization (Adapted with permission from Ref.[41]; Copyright (2016) American Chemical Society)

(1(b)).
[35] NCA

(inimer) NCA.

Lu Cheng^[36] 2007
(HMDS) NCA

(1(c)).
NCA [37].

TMS

[38~40].

C-

(PhS-TMS) NCA

[41] (1(d)).

S—Si

PhS-TMS

NCA

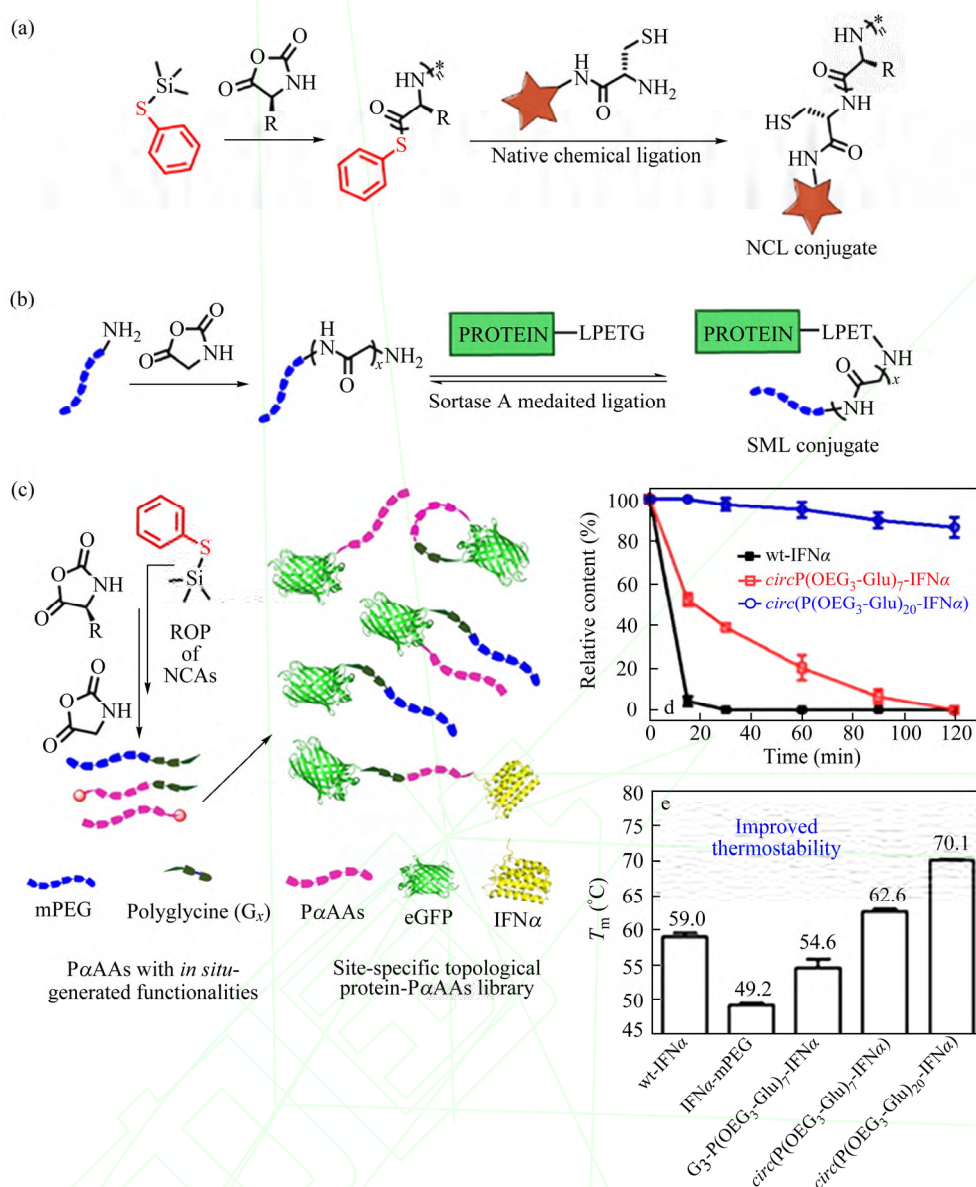


Fig. 2 (a) Synthesis of poly(α -amino acid)s (P α AAs) by PhS-TMS-mediated controlled ring of NCAs (A thioester was installed to the C-terminus of the P α AA and serves as a chemical handle for NCL reaction); (b) A polyglycine (G_x) nucleophile generated by ROP of glycine NCA as a substrate for SML reaction; Combinatorial application of NCL and SML facilely generated various protein-polymer conjugates with novel topology (c), which exhibited excellent protease resistance (d) and thermostability (e) (Adapted with permission from Ref.[44]; Copyright (2016) American Chemical Society)

(IFN-
4
a)
IFN α
IFN α
IFN α
IFN α
(2(d)).
11 °C
[45, 46]
(
2(e)).

4.1

(post-translational
modifications, PTM)

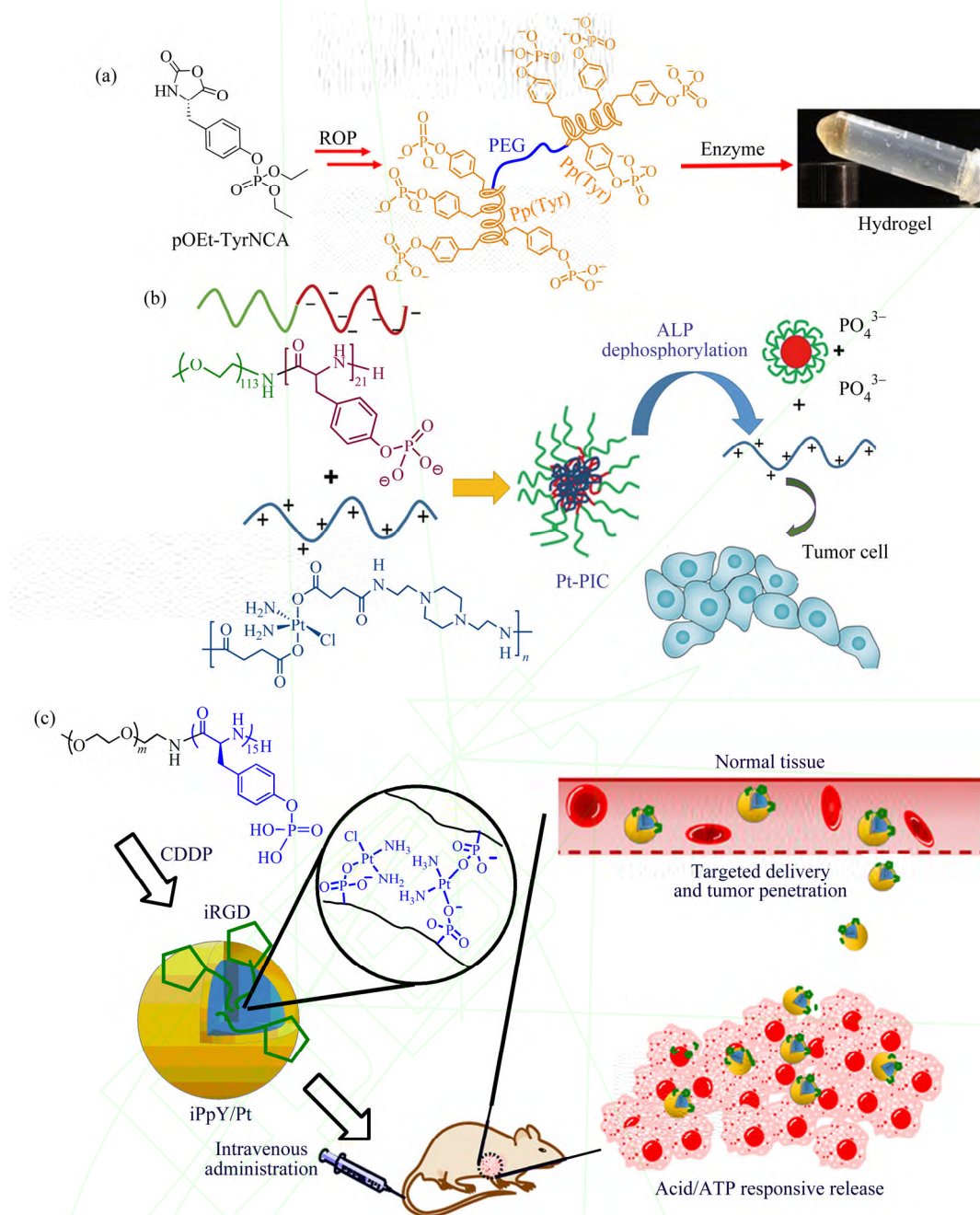


Fig. 3 (a) Synthesis and enzyme-induced hydrogelation of poly(L-phosphotyrosine) (PpY) (Adapted with permission from Ref.[52]; Copyright (2015) American Chemical Society); (b) Schematic illustration of the alkaline phosphatase-triggered dissociation of Pt(IV)-based polyion complex (Pt-PIC) to releases Pt(II) prodrug (Adapted with permission from Ref.[53]; Copyright (2017) The Royal Society of Chemistry); (c) Schematic illustration of acid/ATP-responsive cisplatin release using iPPY/Pt micelle (iPPY/Pt was prepared based on phosphate-patinum complexation and equipped with targeting ligand iRGD) (Adapted with permission from Ref.[54]; Copyright (2017) American Chemical Society)

. Mezzenga [68]

. Yin Cheng [70] 4,5-

-2-

-b-

[69]

NCA

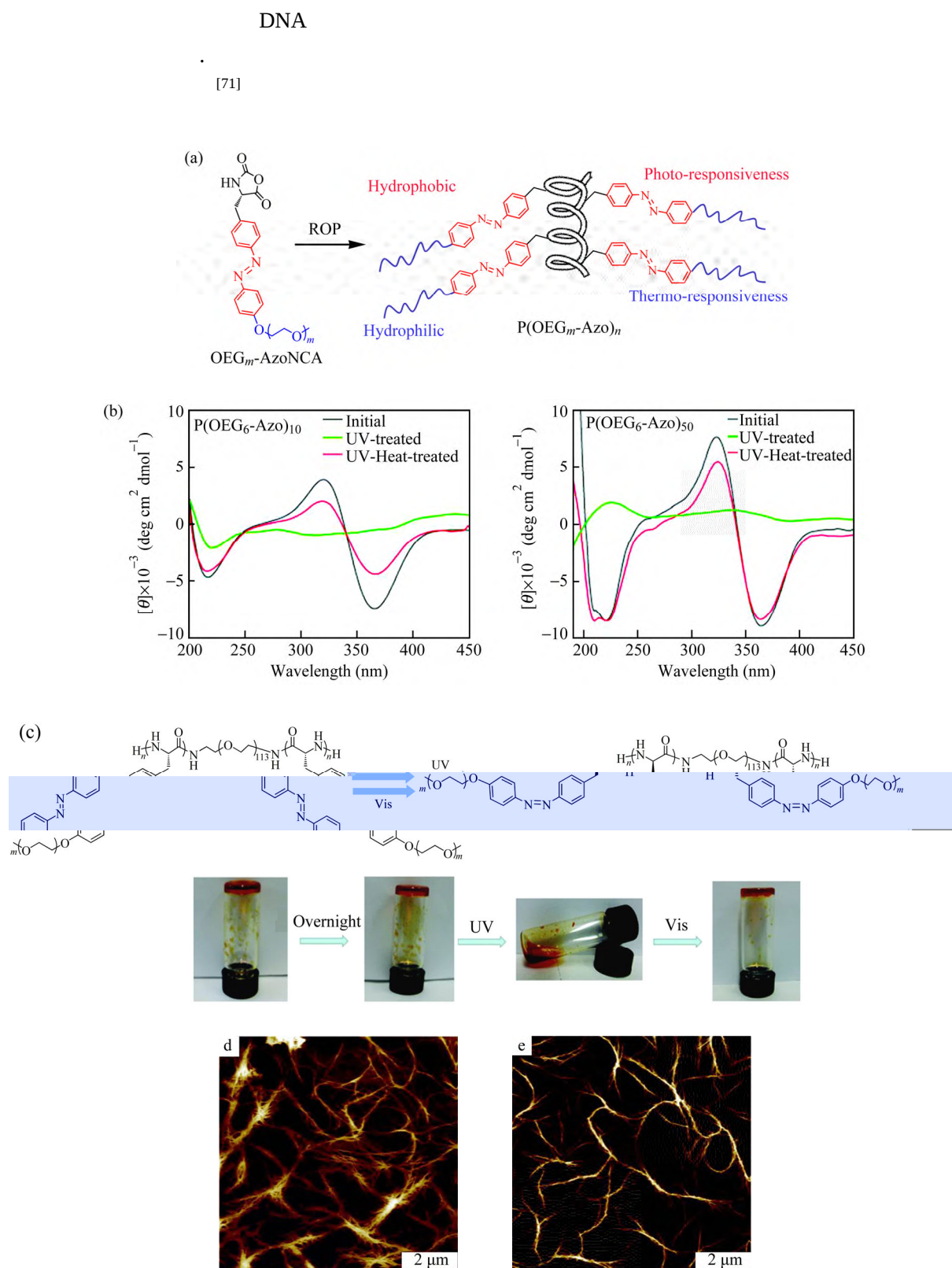
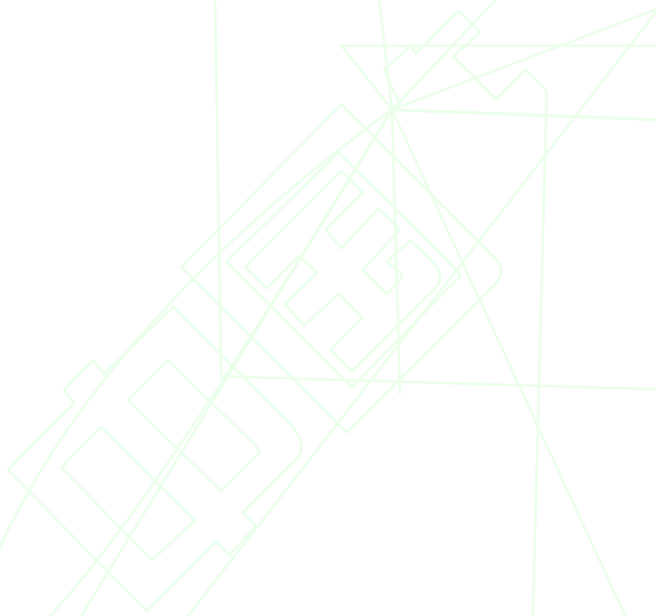


Fig. 4 (a) Synthesis of dual photo- and thermal- responsive P α As; (b) CD spectra of P(OEG₆-Azo)₁₀ and P(OEG₆-Azo)₅₀ in 2,2,2-trifluoroethyl alcohol (the polymers were first irradiated with UV at 365 nm for 5 min, followed by heating at 70 °C for 60 min) (The online version is colorful.); (c) Photographs of triblock copolymer P(OEG₆-Azo)₇-PEG-P(OEG₆-Azo)₇ solution in THF showing reversible UV-Vis triggered gel-sol transition; AFM images of P(OEG₆-Azo)₇-PEG-P(OEG₆-Azo)₇ organogel in its original state (d) and after UV irradiation (e) (Adapted with permission from Ref.[71]; Copyright (2016) The Royal Society of Chemistry)

P(OEG



- 14 Kochendoerfer G G, Chen S Y, Mao F, Cressman S, Traviglia S, Shao H Y, Hunter C L, Low D W, Cagle E N, Carnevali M, Gueriguian V, Keogh P J, Porter H, Stratton S M, Wiedeke M C, Wilken J, Tang J, Levy J J, Miranda L P, Crnogorac M M, Kalbag S, Botti P, Schindler-Horvat J, Savatski L, Adamson J W, Kung A, Kent S B H, Bradburne J A. *Science*, 2003, 299: 884 – 887
- 15 Dumas A, Spicer C D, Gao Z, Takehana T, Lin Y A, Yasukohchi T, Davis B G. *Angew Chem Int Ed*, 2013, 52(14): 3916 – 3921
- 16 Zhang B, Xu H, Chen J, Zheng Y, Wu Y, Si L, Wu L, Zhang C, Xia G, Zhang L, Zhou D. *Acta Biomater*, 2015, 19: 100 – 111
- 17 Biedermann F, Rauwald U, Zayed J M, Scherman O A. *Chem Sci*, 2011, 2(2): 279 – 286
- 18 Cao L, Shi X, Cui Y, Yang W, Chen G, Yuan L, Chen H. *Polym Chem*, 2016, 7(32): 5139 – 5146
- 19 Barz M, Luxenhofer R, Zentel R, Vicent M J. *Polym Chem*, 2011, 2(9): 1900 – 1918
- 20 Merrifield R B. *J Am Chem Soc*, 1963, 85(14): 2149 – 2154
- 21 Tao Youhua(), *Acta Polymerica Sinica*(), 2016, (9): 1151 – 1159
- 22 Huang J, Heise A. *Chem Soc Rev*, 2013, 42(17): 7373 – 7390
- 23 Wang Mingzhi(), Du Jianzhong(). *Acta Polymerica Sinica*(), 2014, (9): 1183 – 1194
- 24 Cheng J, Deming T J. *Topics in Current Chemistry*. Springer Berlin Heidelberg, 2012, 310: 1 – 26
- 25 Leuchs H. *Chem Ber*, 1906, 39: 857 – 861
- 26 Deming T J. *Nature*, 1997, 390(6658): 386 – 389
- 27 Deming T J. *J Am Chem Soc*, 1998, 120(17): 4240 – 4241
- 28 Dimitrov I, Schlaad H. *Chem Commun*, 2003, 23: 2944 – 2945
- 29 Conejos-Sánchez I, Duro-Castano A, Birke A, Barz M, Vicent M J. *Polym Chem*, 2013, 4(11): 3182 – 3186
- 30 Peng Y, Lai S, Lin C. *Macromolecules*, 2008, 41(10): 3455 – 3459
- 31 Peng H, Ling J, Shen Z. *J Polym Sci, Part A: Polym Chem*, 2012, 50(6): 1076 – 1085
- 32 Aliferis T, Iatrou H, Hadjichristidis N. *Biomacromolecules*, 2004, 5(5): 1653 – 1656
- 33 Zhao W, Gnanou Y, Hadjichristidis N. *Biomacromolecules*, 2015, 16(4): 1352 – 1357
- 34 Zhao W, Gnanou Y, Hadjichristidis N. *Polym Chem*, 2015, 6(34): 6193 – 6201
- 35 Li P, Dong C. *ACS Macro Lett*, 2017, 6(3): 292 – 297
- 36 Lu H, Cheng J. *J Am Chem Soc*, 2007, 129(46): 14114 – 14115
- 37 Lu H, Cheng J. *J Am Chem Soc*, 2008, 130(38): 12562 – 12563
- 38 Lu H, Wang J, Lin Y, Cheng J. *J Am Chem Soc*, 2009, 131(38): 13582 – 13583
- 39 Bai Y, Lu H, Ponnusamy E, Cheng J. *Chem Commun*, 2011, 47(38): 10830 – 10832
- 40 Baumgartner R, Fu H, Song Z, Lin Y, Cheng J. *Nat Chem*, 2017, 9(7): 614 – 622
- 41 Yuan J, Sun Y, Wang J, Lu H. *Biomacromolecules*, 2016, 17(3): 891 – 896
- 42 Grotzky A, Manaka Y, Kojima T, Walde P. *Biomacromolecules*, 2010, 12(1): 134144
- 43 Talelli M, Vicent M J. *Biomacromolecules*, 2014, 15(11): 4168 – 4177
- 44 Hou Y, Yuan J, Zhou Y, Yu J, Lu H. *J Am Chem Soc*, 2016, 138(34): 10995 – 11000
- 45 Deng C, Wu J, Cheng R, Meng F, Klok H A, Zhong Z. *Prog Polym Sci*, 2014, 39(2): 330 – 364
- 46 Lu H, Wang J, Song Z, Yin L, Zhang Y, Tang H, Tu C, Lin Y, Cheng J. *Chem Commun*, 2014, 50(2): 139 – 155
- 47 Walsh C T, Garneau-Tsodikova S, Gatto G J. *Angew Chem Int Ed*, 2005, 44(45): 7342 – 7372
- 48 Kramer J R, Deming T J. *Polym Chem*, 2014, 5(3): 671 – 682
- 49 Pratt M R, Bertozzi C R. *Chem Soc Rev*, 2005, 34(1): 58 – 68
- 50 Pati D, Shaikh A Y, Das S, Nareddy P K, Swamy M J, Hotha S, Gupta S S. *Biomacromolecules*, 2012, 13(5): 1287 – 1295
- 51 Rubin C S, Rosen O M. *Annu Rev Biochem*, 1975, 44(1): 831 – 887
- 52 Sun Y, Hou Y, Zhou X, Yuan J, Wang J, Lu H. *ACS Macro Lett*, 2015, 4(9): 1000 – 1003
- 53 Li S, Hou Y, Hu Y, Yu J, Wei W, Lu H. *Biomater Sci*, 2017, 5: 1558 – 1566
- 54 Hou Y, Wang Y, Wang R, Bao W, Xi X, Sun Y, Lu H. *ACS Appl Mater Interfaces*, 2017, 9(21): 17757 – 17768
- 55 Shen Y, Fu X, Fu W, Li Z. *Chem Soc Rev*, 2015, 44(3): 612 – 622
- 56 Yuan Jingsong(), Lu Hua(). *Polym Bull*(), 2015, (9): 210 – 216
- 57 Chen C, Wang Z, Li Z. *Biomacromolecules*, 2011, 12(8): 2859 – 2863
- 58 Fu X, Ma Y, Sun J, Li Z. *Chinese J Polym Sci*, 2016, 34(12): 1436 – 1447
- 59 Zhang S, Chen C, Li Z. *Chinese J Polym Sci*, 2013, 2(31): 201 – 210

- 60 Ge C, Liu S, Liang C, Ling Y, Tang H. *Polym Chem*, 2016, 7(38): 5978 – 5987
- 61 Shi F, Ding J, Xiao C, Zhuang X, He C, Chen L, Chen X. *J Mater Chem*, 2012, 22(28): 14168 – 14179
- 62 Wang K, Luo G, Liu Y, Li C, Cheng S, Zhuo R, Zhang X. *Polym Chem*, 2012, 3(4): 1084 – 1090
- 63 Kramer J R, Deming T J. *J Am Chem Soc*, 2012, 134(9): 4112 – 4115
- 64 Chen P, Qiu M, Deng C, Meng F, Zhang J, Cheng R, Zhong Z. *Biomacromolecules*, 2015, 16(4): 1322 – 1330
- 65 Lee E S, Oh K T, Kim D, Youn Y S, Bae Y H. *J Control Release*, 2007, 123(1): 19 – 26
- 66 Rodríguez-Hernández J, Lecommandoux S. *J Am Chem Soc*, 2005, 127(7): 2026 – 2027
- 67 Bae Y, Fukushima S, Harada A, Kataoka K. *Angew Chem Int Ed*, 2003, 42(38): 4640 – 4643
- 68 Kotharangannagari V K, Sánchez-Ferrer A, Ruokolainen J, Mezzenga R. *Macromolecules*, 2011, 44(12): 4569 – 4573
- 69 Liu G, Dong C. *Biomacromolecules*, 2012, 13(5): 1573 – 1583
- 70 Yin L, Tang H, Kim K H, Zheng N, Song Z, Gabrielson N P, Lu H, Cheng J. *Angew Chem Int Ed*, 2013, 52(35): 9182 – 9186
- 71 Xiong W, Fu X, Wan Y, Sun Y, Li Z, Lu H. *Polym Chem*, 2016, 7(41): 6375 – 6382

Efficient Synthesis and Application of Protein-Poly(α -amino acid) Conjugates

Chong Zhang, Hua Lu*

(Beijing National Laboratory for Molecular Sciences, Key Laboratory of Polymer Chemistry and Physics of Ministry of Education, Center for Soft Matter Science and Engineering, College of Chemistry and Molecular Engineering, Peking University, Beijing 100871)

Abstract Protein-polymer conjugates are important therapeutics for various diseases. There are currently two major challenges in this field: one is the search of new biodegradable polymers beyond traditional PEGylation, and the other is to develop highly efficient and site-specific conjugation strategy. Poly(α -amino acid)s (PaAAs) are biodegradable and biocompatible polymers with tunable properties and numerous functions, making them promising candidates for protein modification. In this review, we summarize our recent progresses in protein-PaAAs conjugates. Specifically, we discuss our developments in: (1) Recent developments in the controlled ring-opening polymerization (ROP) of α -amino acid *N*-carboxyanhydrides (NCAs), including amine-based initiators, organometallic initiators, organosilicon amines initiators and sulfide-based initiators. For instance, trimethylsilyl phenylsulfide (PhSTMS) is a novel initiator for controlled ROP of NCAs. It exhibits higher nucleophilicity than conventional amine-based initiator, and thus affords considerably higher chain initiation rate to ensure a more controlled polymerization. Moreover, this initiator is well-tolerated to various functional groups. (2) *In situ* functionalization of PaAAs for site-specific protein conjugation, and construction of various topological structures. Using PhSTMS initiator, it *in situ* generates a reactive phenyl thioester group at one end of the PaAAs, which can be used for protein *N*-terminus conjugation *via* native chemical ligation (NCL); moreover, ROP of glycine NCA yields oligoglycine at the other end of PaAAs, which can be used for C-terminus protein conjugation *via* sortase-A mediated ligation (SML). More interestingly, combinatory use of the two methods can construct various topological protein-PaAA conjugates including the head-to-tail circular conjugates. (3) Development of functional PaAAs for potential protein conjugation. Various functional PaAAs have been developed as delivery materials or hydrogels. To further expand the arsenal of PaAAs for potential modulation of protein functions, PaAAs that mimic protein post-translational modifications (PTM) are synthesized; On the other hand, a series of multiple stimuli-responsive PaAAs are also produced. These PaAAs show interesting enzyme, light, and/or thermal responsiveness, which could be potentially harnessed for modulation of protein functions in the future.

Keywords Protein conjugation, Poly(α -amino acid), PTM mimicking, Topology, Stimuli-responsiveness

* Corresponding author: Hua Lu, E-mail: chemhualu@pku.edu.cn